

基础研究

右美托咪啶预处理对脓毒症肾损伤大鼠炎症因子和氧化应激的影响

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摘要:目的 探讨右美托咪啶对脓毒症大鼠AKI(急性肾损伤)的炎症因子和氧化应激的影响。方法 将32只雄性SD大鼠随机均分为以下4组(每组8只大鼠):假手术组、脂多糖(LPS)组、右美托咪啶(DEX)+LPS组、育亨宾(YOH)+DEX+LPS组;后3组分别于术前30 min经尾静脉注射LPS(5 mg/kg);LPS + DEX组LPS前10 min尾静脉注射DEX(10 μ g/kg);YOH+DEX+LPS组于LPS前40 min经腹腔注射YOH(1 mg/kg),及LPS前10min尾静脉注射DEX 10 μ g/kg。4 h后处死大鼠提取标本测定血浆和肾组织中的白介素-1 β (IL-1 β)、超氧化物歧化酶(SOD)和丙二醛(MDA)水平,并观察肾组织病理学变化。结果 与假手术组相比,LPS组中血浆和肾组织IL-1 β 、MDA水平明显升高,SOD明显降低($P<0.05$),肾脏病理损伤严重;与LPS组相比,DEX +LPS组中血浆和肾组织IL-1 β 、MDA水平明显降低,SOD明显增高($P<0.05$),肾脏病理学损伤也明显减轻;YOH+DEX+LPS组和DEX +LPS组相比,IL-1 β 、MDA均上升,SOD下降($P<0.05$),肾脏病理学损伤较明显。结论 DEX可以减轻脓毒症相关肾损伤的炎症反应和氧化应激,且这种作用可能是通过 α_2 受体起作用的。

关键词:右美托咪啶; α_2 肾上腺受体;脓毒症肾损伤;氧化应激;炎症反应

Pretreatment with dexmedetomidine ameliorates renal inflammation and oxidative stress in rats with lipopolysaccharide-induced sepsis and acute kidney injury

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Abstract: **Objective** To investigate the effects of dexmedetomidine on inflammatory reaction, oxidative stress, and renal pathologies in a rat model of lipopolysaccharide (LPS)-induced sepsis. **Methods** Thirty-two SD rats were randomly divided into 4 groups, including a sham-operated group, LPS group with LPS (5 mg/kg) injection via the caudal vein 30 min before the operation, dexmedetomidine (Dex) +LPS group with additional Dex (10 μ g/kg) injection via the caudal vein 10 min before LPS injection, and yohimbine+DEX+LPS group with intraperitoneal yohimbine (1 mg/kg) injection 40 min before and Dex injection 10 min before LPS injection. The levels of IL-1 β , SOD and MDA in the plasma and renal tissues were determined, and the renal pathologies were examined. **Results** Compared with the sham-operated rats, the rats in LPS group showed significantly increased IL-1 β and MDA levels and lowered SOD activity in the plasma and renal tissues ($P<0.05$) with obvious renal pathologies. Dex pretreatment obviously lowered IL-1 β and MDA levels and enhanced SOD activity in the plasma and renal tissues in LPS-challenged rats ($P<0.05$), and significantly lessened LPS-induced renal pathologies. **Conclusion** Dex can protect the rats against LPS-induced renal injury by alleviating the inflammatory reactions and cytokine oxidative stress, and this effect is mediated possibly by α_2 receptors.

Key words: dexmedetomidine; α_2 adrenoceptor; sepsis; renal damage; oxidative stress; inflammatory reaction

脓毒症,是由于各种病原体感染、创伤、烧伤、缺氧、再灌注损伤及外科大手术后引起的一个严重的、复杂的、失控的、全身性的炎症反应综合症,可引起感染性休克及多器官功能障碍综合征。肾脏是最易受到脓毒症打击的靶器官之一,脓毒症患者中约42%发生急性肾损伤(acute kidney injury, AKI)^[1],研究表明约50%的重

症AKI患者的病因主要是脓毒症,尽管使用抗生素或免疫调节治疗应用,仍然不能有效地控制病人的多器官功能损伤和病人死亡,由脓毒症引发AKI导致患者死亡率仍然居高不下,脓毒症患者发生AKI后死亡率高达70%^[2-3]。而ICU里需要透析治疗的AKI病人生存率仅有13.8%。因此,为了降低脓毒症导致AKI发病率和改善预后,如何及时预防和有效干预脓毒症并发AKI成为目前一项迫切研究课题。

目前众多研究证实机体脓毒症可导致AKI甚至多器官功能障碍综合征,可能与机体交感神经过度兴奋、

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免疫失调、炎症反应失控等一系列病理生理改变有关。因此,在脓毒症的治疗中,抑制交感神经过度兴奋和炎症级联放大效应,减轻脓毒症导致AKI是降低脓毒症病死率的有效措施,也是当今危重症医学领域研究的热点和难点。

右美托咪啶(DEX)是选择性 α_2 -肾上腺受体激动剂,对位于脑和脊髓的 α_2 肾上腺素能受体(α_2 -AR)产生激动效应,产生镇静、镇痛和抑制交感神经等作用,临床上主要用于镇静、镇痛、抑制交感活动等治疗。近年来,越来越多的基础和临床研究证明,DEX能抑制中性粒细胞的激活及TNF- α 、IL-1 β 等炎性细胞因子的释放而发挥全身的抗炎作用,另外也可抑制致炎因子导致的中性粒细胞的呼吸爆发,避免脂质过氧化反应,对心、肾、肺等多器官具有保护作用^[4-6]。本研究旨在探讨DEX对脓毒症大鼠AKI模型中肾脏组织细胞的影响,以及炎性因子和氧化应激反应扮演的具体作用机制,希望为AKI的临床治疗提供理论依据。

1 材料与方法

1.1 实验动物和材料

雄性清洁级SD大鼠32只,体质量200~250 g(中山大学动物实验中心提供)。右美托咪啶(规格:200 μ g/mL,江苏恒瑞制药厂),育亨宾(yohimbine, YOH)(Sigma-Aldrich),IL-1 β 试剂盒(南京凯基生物试剂有限公司),MDA和SOD试剂盒(南京建成生物工程研究所),脂多糖(Sigma-Aldrich)。

1.2 动物分组和模型制备

大鼠随机均分为以下4组:Sham组、LPS组、DEX+LPS组和YOH+DEX+LPS组。采用LPS诱导制备脓毒症血症肾损伤模型,大鼠常规禁食12 h,不禁饮。诱导麻醉大鼠后,将其放置于变温毯上仰卧,固定,维持体温 36 ± 0.1 $^{\circ}\text{C}$ 。除育亨宾经腹腔注射外,DEX和LPS均通过尾静脉注射。具体给药方式如下:LPS组给予LPS 5 mg/kg;DEX+LPS组,首先注射DEX 10 μ g/kg,10 min后同样给予LPS 5 mg/kg;YOH+DEX+LPS组,先经腹腔注射YOH 1 mg/kg,30 min后注射DEX 10 μ g/kg,10 min后给予LPS 5 mg/kg。Sham组给予与LPS等量生理盐水注射。

1.3 标本的留取

4 h后打开腹腔,经腹主动脉抽取血液,评估IL-1 β 、SOD及MDA检测。截取左肾脏取部分组织,用10%福尔马林固定,石蜡包埋,切片,行HE染色,光镜下进行组织病理学评估;右肾组织迅速放入液氮中保存,实验结束后转入 -80 $^{\circ}\text{C}$ 低温冰箱保存,用于肾组织匀浆检测IL-1 β 、SOD和MDA水平。

1.4 MDA、SOD和IL-1 β 检测

血和肾组织采用黄嘌呤氧化酶法检测SOD的含量

和活性,用硫代巴比妥酶法测定MDA的含量,具体按照操作试剂盒说明书进行,肾组织中的SOD、MDA含量按照公式计算,考马斯亮蓝检测蛋白含量。IL-1 β 采用酶联免疫分析法(ELISA)操作试剂盒说明书进行。

1.5 统计学分析

所得计量数据均以GraphPad Prism 5软件统计,组内比较采用重复测量方差分析,组间比较采用 t 检验 $P<0.05$ 为差异有统计学意义。

2 结果

2.1 右美托咪啶预处理对血浆和肾组织MDA和SOD的影响

2.1.1 各组血浆和肾组织MDA水平变化 与Sham组相比,LPS组明显增加血浆和肾脏的MDA含量($P<0.05$);与LPS组相比,右美托咪啶预处理组(DEX+LPS组)明显降低血浆和肾脏的MDA含量($P<0.05$);与DEX+LPS组相比, α_2 -肾上腺素受体拮抗剂育亨宾组(YOH+DEX+LPS组)MDA含量上升($P<0.05$,图1)。

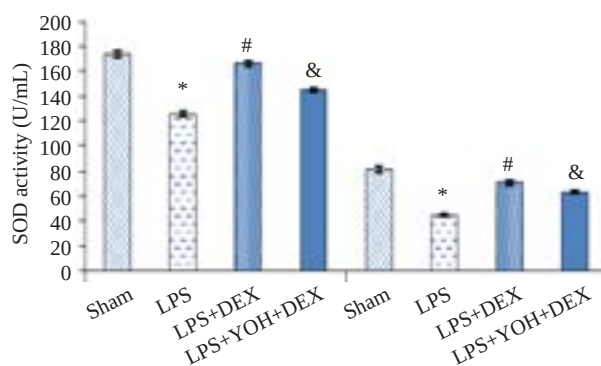


图1 各组血浆和肾组织MDA水平的变化

Fig.1 Levels of MDA in the blood and kidney tissue in different groups. Compared with Sham group, LPS group * $P<0.05$; Compared with LPS group, DEX+LPS group # $P<0.05$; Compared with DEX+LPS group, YOH+DEX+LPS group & $P<0.05$.

2.1.2 各组血浆和肾组织SOD活力变化 与Sham组相比,LPS组明显降低血浆和肾脏的SOD活力($P<0.05$);与LPS组相比,DEX+LPS组明显升高血浆和肾脏的SOD活力($P<0.05$);与DEX+LPS组相比,YOH+DEX+LPS组SOD活力下降($P<0.05$,图2)。

2.2 右美托咪啶预处理对血浆和肾组织IL-1 β 的影响

与Sham组相比,LPS组明显增加血浆和肾脏的IL-1 β 含量($P<0.05$);与LPS组相比,DEX+LPS组明显降低血浆和肾脏的IL-1 β 含量($P<0.05$);与DEX+LPS组相比,YOH+DEX+LPS组IL-1 β 含量上升($P<0.05$,图3)。

2.3 肾脏病理学改变

Sham组肾组织结构正常,肾小管、肾间质未见明显

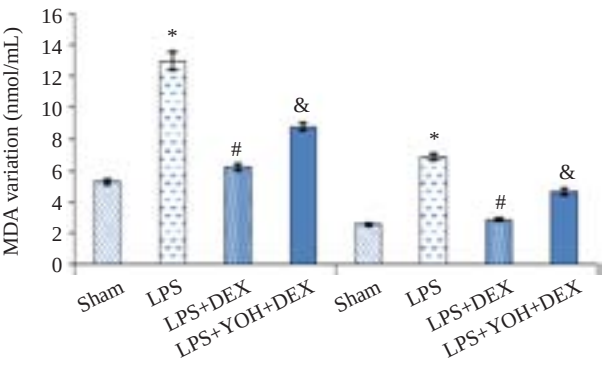


图2 各组血浆和肾组织SOD活力的变化

Fig.2 SOD activity in the blood and kidney tissue in different groups. Compared with Sham group, LPS group * $P<0.05$; Compared with LPS group, DEX+LPS group # $P<0.05$; Compared with DEX+LPS group ,YOH+DEX+LPS group & $P<0.05$.

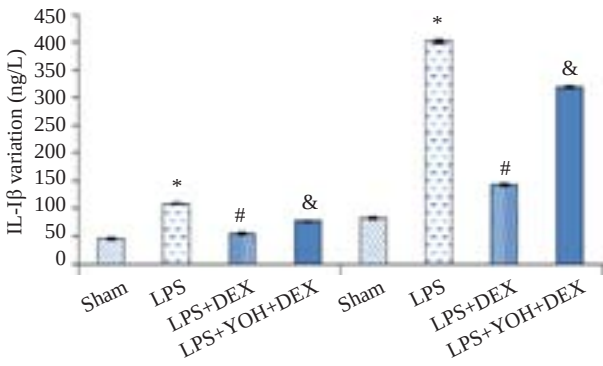


图3 各组血浆和肾组织IL-1β水平的变化

Fig.3 Levels of IL-1β in the blood and kidney tissue in different groups. Compared with Sham group, LPS group * $P<0.05$; Compared with LPS group, DEX+LPS group # $P<0.05$; Compared with DEX+LPS group ,YOH+DEX+LPS group & $P<0.05$.

的病理性改变;LPS组肾小管上皮细胞肿胀,部分出现空泡变性,肾间质炎症细胞浸润;DEX+LPS组肾脏组织病理损伤减轻,肾小管上皮细胞轻度肿胀或扁平,少量出现空泡样变,肾间质炎症细胞浸润减少;YOH+DEX+LPS组肾脏组织病理损伤较DEX+LPS组加重(图4)。

3 讨论

研究发现,DEX预处理大鼠术后4 h,病理学检查发现DEX明显减轻大鼠肾小管上皮细胞变性,肾小管囊腔扩张、空泡变性,肾间质炎症细胞浸润,证明DEX预处理可以显著减轻LPS诱导产生的肾小管损伤。近

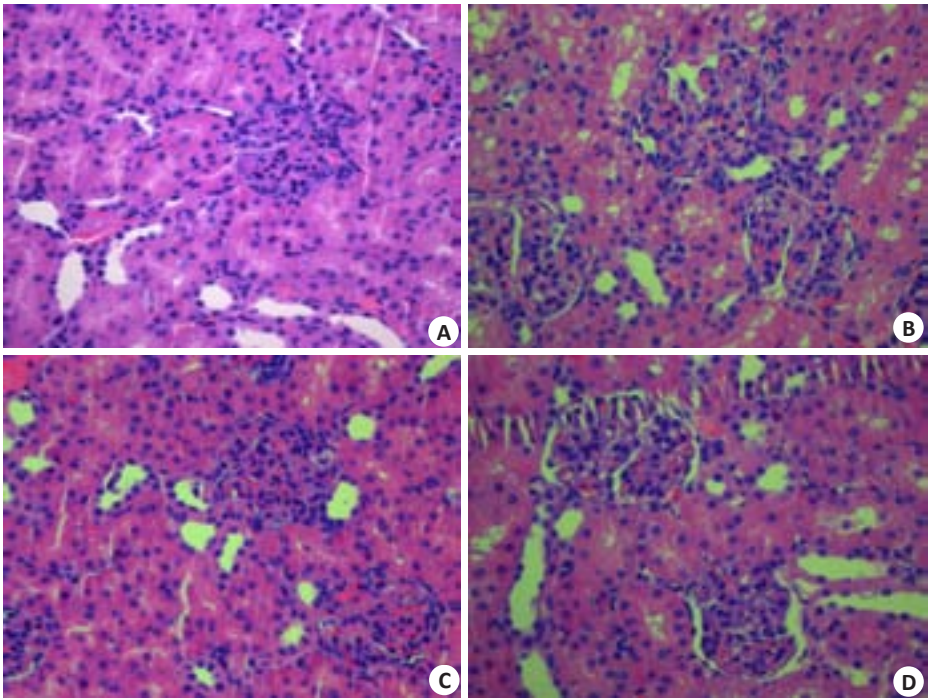


图4 各组肾脏组织的病理学改变

Fig.4 Histopathological changes in the kidney tissue in different groups (HE staining, original magnification: $\times 400$). A: SHAM group; B: LPS group; C: DEX+LPS group; D: YOH+DEX+LPS group.

年来国内外学者研究发现DEX对脑、心、肝、肺、和小肠等多个器官具有保护作用,认为其保护机制是与抗炎、抗氧化损伤与抑制细胞凋亡有关^[7-11],本实验通过采用LPS诱导制备大鼠脓毒症AKI模型,应用DEX显著减

轻脓毒症大鼠氧化应激和炎症反应,而这一保护作用被DEX特异性受体拮抗剂育亨宾逆转,推测DEX的AKI保护机制可能是通过肾小管上皮细胞上的α2受体起保护作用。

AKI是十分复杂的病理生理过程,其发病机制可能涉及肾缺血再灌注损伤、炎症因子、一氧化氮学说、细胞凋亡学说、内皮功能障碍及内皮素作用、内毒素直接损伤、氧化应激损伤等。LPS相关的AKI与炎症和肾小管的损伤密切相关, $\alpha 2$ 肾上腺素能受体广泛分布于全身多种器官、组织和细胞,在肾脏, $\alpha 2$ 受体主要分布于肾近曲小管、集合管及微血管等。DEX是新一代高选择性 $\alpha 2$ 肾上腺素能受体激动剂^[12]。DEX具有抗交感兴奋、调节免疫功能、氧化应激及抗炎反应等作用特点,在脓毒症的治疗及器官保护方面具有显著优势^[5, 13-14]。近年来有不少关于DEX对肾脏保护作用机制的研究,认为DEX对脓毒症肾损伤的保护机制主要与抗氧化应激、抗炎反应及抗细胞凋亡等有关。如Villela等^[15]发现DEX可以通过减少肾脏交感神经作用,降低尿渗透压和血浆精氨酸加压素水平,产生利尿作用。Gu等^[16]研究指出DEX通过激动 $\alpha 2$ AR,抑制肾脏细胞凋亡,下调TLR4蛋白表达以及血清HMGB1的水平,是其肾保护的主要机制。另外DEX通过激动 $\alpha 2$ 受体抑制多条信号通路如JAK/STAT信号通路、TLR-4/NF-KB/MAPKs通路下调炎症产物发挥抗炎作用^[17-19]。本实验检测了血中的IL-1 β 水平,发现LPS注射诱导内毒素AKI后IL-1 β 水平明显增高,DEX可以下调炎症因子IL-1 β 的表达,该作用被育亨宾部分拮抗。本实验检测结果和国内外学者研究结论是相符的^[5, 20-22]。

内毒素血症导致线粒体内呼吸链产生的活性氧簇增加,氧自由基产生增多;另外肾小管上皮细胞内酶活性降低,导致机体产生大量的氧自由基,攻击生物膜中的不饱和脂肪酸,加速脂质过氧化过程,脂质过氧化产物如MDA产生增加,肾小管上皮细胞损伤和功能代谢障碍,甚至产生细胞凋亡,这也是内毒素诱发的肾小管上皮细胞损伤的重要机制之一。我们的结果显示,内毒素血症导致肾组织和血浆中MDA含量明显升高,SOD活力明显下降,表明内毒素注射后机体和肾组织中的氧自由基增加,脂质过氧化程度加剧,抗氧化酶活力下降。DEX预处理能明显降低内毒素血症中肾组织和血浆中的MDA含量,增加SOD活力,提示DEX介导的SOD水平提高可能有助于减少内毒素导致AKI产生的氧自由基,减轻氧化应激,发挥内毒素AKI的保护作用,和国内外的研究一致^[23-26],而 $\alpha 2$ -肾上腺素受体拮抗剂YOH能部分逆转SOD和MDA值的变化。

本实验采用LPS诱导制备大鼠脓毒症模型,这是目前公认的与临床相关性较强的脓毒症模型,证明了DEX可以明显减轻LPS诱导的急性肾损伤的氧化应激和炎症反应,减轻了肾脏的病理损伤程度, $\alpha 2$ -肾上腺素受体拮抗剂YOH能部分拮抗此作用,可以推断DEX对内毒素肾损伤的保护作用可能和免疫炎症调节和抗氧

化产物生成密切相关。但其调节信号通路中的具体作用机制尚不清楚,有待于进一步实验探索。

参考文献:

- [1] Bagshaw SM, George C, Bellomo R, et al. Early acute kidney injury and sepsis: a multicentre evaluation[J]. Crit Care, 2008, 12(2): R47.
- [2] Mehta RL, Bouchard J, Soroko SB, et al. Sepsis as a cause and consequence of acute kidney injury: program to improve care in acute renal disease[J]. Intensive Care Med, 2011, 37(2): 241-8.
- [3] Rajapakse S, Rodrigo C, Rajapakse A, et al. Renal replacement therapy in sepsis induced acute renal failure[J]. Saudi J Kidney Dis Transpl, 2009, 20(4): 553-9.
- [4] Sanders RD, Sun P, Patel S, et al. Dexmedetomidine provides cortical neuroprotection: impact on anaesthetic-induced neuroapoptosis in the rat developing brain [J]. Acta Anaesthesiol Scand, 2010, 54(6): 710-6.
- [5] Taniguchi T, Kurita A, Kobayashi K, et al. Dose- and time-related effects of dexmedetomidine on mortality and inflammatory responses to endotoxin-induced shock in rats[J]. J Anesth, 2008, 22(3): 221-8.
- [6] Hofer S, Steppan J, Wagner T, et al. Central sympatholytics prolong survival in experimental sepsis[J]. Crit Care, 2009, 13(1): R11.
- [7] Riker RR, Shehabi Y, Bokesch PM, et al. Dexmedetomidine vs midazolam for sedation of critically ill patients: a randomized trial [J]. JAMA, 2009, 301(5): 489-99.
- [8] Mantz J, Josserand J, Hamada S. Dexmedetomidine: new insights [J]. Eur J Anaesthesiol, 2011, 28(1): 3-6.
- [9] Hanci V, Yurdakan G, Yurtlu S, et al. Protective effect of dexmedetomidine in a rat model of alpha-naphthylthiourea-induced acute lung injury[J]. J Surg Res, 2012, 178(1): 424-30.
- [10] Gu J, Chen J, Xia P, et al. Dexmedetomidine attenuates remote lung injury induced by renal ischemia-reperfusion in mice [J]. Acta Anaesthesiol Scand, 2011, 55(10): 1272-8.
- [11] Gazit AZ. Dexmedetomidine and pulmonary artery pressure in the pediatric cardiac surgery patient-A new insight[J]. Pediatr Crit Care Med, 2010, 11(5): 634-5.
- [12] Kaur M, Singh PM. Current role of dexmedetomidine in clinical anesthesia and intensive care[J]. Anesth Essays and Res, 2011, 5(2): 128-33.
- [13] Hardin KA. Sleep in the ICU potential mechanisms and clinical implications[J]. Chest, 2009, 136(1): 284-94.
- [14] Lin Y, Zhu X, Yao WZ, et al. Yohimbine protects against endotoxin-induced acute lung injury by blockade of alpha 2A adrenergic receptor in rats [J]. Chin Med J (Engl), 2011, 124(7): 1069-74.
- [15] Villela NR, Do Nascimento JP, Carvalho LR, et al. Effects of dexmedetomidine on renal system and on vasopressin plasma levels. Experimental study in dogs[J]. Rev Bras Anesthesiol, 2005, 55(4): 429-40.
- [16] Gu J, Chen J, Xia P, et al. Dexmedetomidine attenuates remote lung injury induced by renal ischemia-reperfusion in mice [J]. Acta Anaesthesiol Scand, 2011, 55(10): 1272-8.

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- [9] 吴韶清, 廖 灿, 黄以宁, 等. 广州地区6493例女性生殖道人乳头状瘤病毒检测结果分析[J]. 中国实用妇科与产科杂志, 2011(6): 453-6.
- [10] 庾永基, 廖日房, 吴北湖, 等. 广州市妇女人乳头瘤病毒感染型别的检测[J]. 中国卫生检验杂志, 2013(15): 3072-4.
- [11] 李发涛, 廖 灿, 李 焱, 等. 人乳头状瘤病毒感染妇女的年龄及其亚型分析[J]. 中华医院感染学杂志, 2014(17): 4345-7.
- [12] Liu SS, Chan KY, Leung RC, et al. Prevalence and risk factors of human papillomavirus (HPV) infection in southern Chinese women-a population-based study[J]. PLoS One, 2011, 6(5): e19244.
- [13] 林 丹, 蒋惠萍, 张 易, 等. 广州社区女性宫颈HPV感染相关因素调查[J]. 广东医学, 2013(16): 2563-5.
- [14] Jing L, Zhong X, Zhong Z, et al. Prevalence of human papillomavirus infection in Guangdong Province, China: a population-based survey of 78, 355 women[J]. Sex Transm Dis, 2014, 41(12): 732-8.
- [15] 林 盈, 姜晓琦, 严 英. 深圳龙岗地区宫颈人乳头瘤病毒感染流行病学调查[J]. 中外医学研究, 2014(23): 68-9.
- [16] 黄玉芳, 张 玲, 王厚照. 厦门地区女性人乳头瘤病毒感染状况调查[J]. 国际病毒学杂志, 2014, 21(4): 181-5.
- [17] 钱慧珍, 范晓明, 张 君, 等. 大连地区女性人乳头瘤病毒感染状况及基因分型分布情况调查[J]. 中国医师进修杂志, 2014, 37(z1): 29-31.
- [18] Ghittoni R, Accardi R, Chiocca S, et al. Role of human papillomaviruses in carcinogenesis [J]. Ecancermedalscience, 2015, 9: 526.
- [19] Dickson EL, Vogel RI, Geller MA, et al. Cervical cytology and multiple type HPV infection: a study of 8182 women ages 31-65[J]. Gynecol Oncol, 2014, 133(3): 405-8.
- [20] Chaturvedi AK, Katki HA, Hildesheim A, et al. human papillomavirus infection with multiple types: pattern of coinfection and risk of cervical disease[J]. J Infect Dis, 2011, 203(7): 910-20.
- [21] Spinillo A, Gardella B, Roccio M, et al. Multiple human papillomavirus infection with or without type 16 and risk of cervical intraepithelial neoplasia among women with cervical cytological abnormalities[J]. Cancer Causes Control, 2014, 25(12): 1669-76.
- [22] Smith JS, Lindsay L, Hoots B, et al. Human papillomavirus type distribution in invasive cervical cancer and high-grade cervical lesions: a meta-analysis update [J]. Int J Cancer, 2007, 121(3): 621-32.
- [23] Wu Y, Chen Y, Li L, et al. Associations of high-risk HPV types and viral load with cervical cancer in China[J]. J Clin Virol, 2006, 35 (3): 264-9.
- [24] Shen Y, Gong JM, Li YQ, et al. Epidemiology and genotype distribution of human papillomavirus (HPV) in women of Henan Province, China[J]. Clin Chim Acta, 2013, 415: 297-301.
- [25] Mariani L, Vici P, Suligoi B, et al. Early direct and indirect impact of quadrivalent HPV (4HPV) vaccine on genital warts: a systematic review[J]. Adv Ther, 2015, 32(1): 10-30.
- [26] 宋云焕, 周自广. 二价HPV疫苗预防宫颈癌及HPV相关感染的meta分析[J]. 山西医科大学学报, 2012, 43(5): 377-81.
- (编辑:吴锦雅)

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- [17] Nance DM, Sanders VM. Autonomic innervation and regulation of the immune system (1987-2007) [J]. Brain Behav Immun, 2007, 21 (6): 736-45.
- [18] Si Y, Bao H, Han L, et al. Dexmedetomidine protects against renal ischemia and reperfusion injury by inhibiting the JAK/STAT signaling activation[J]. J Transl Med, 2013, 11(1): 141.
- [19] Lai YC, Tsai PS, Huang CJ. Effects of dexmedetomidine on regulating endotoxin-induced up-regulation of inflammatory molecules in murine macrophages [J]. J Surg Res, 2009, 154(2): 212-9.
- [20] Hsing CH, Lin CF, So E, et al.  $\alpha$ 2-A adrenoreceptor agonist dexmedetomidine protects septic acute kidney injury through increasing BMP-7 and inhibiting HDAC2 and HDAC5 [J]. Am J Physiol Renal Physiol, 2012, 303(10): F1443-53.
- [21] Yang CL, Tsai PS, Huang CJ. Effects of dexmedetomidine on regulating pulmonary inflammation in a rat model of ventilator-induced lung injury [J]. Acta Anaesthesiol Taiwan, 2008, 46(4): 151-9.
- [22] Tasdogan M, Memis D, Sut N, et al. Results of a pilot study on the effects of propofol and dexmedetomidine on inflammatory responses and intraabdominal pressure in severe sepsis [J]. J Clin Anesth, 2009, 21(6): 394-400.
- [23] Eser O, Fidan H, Sahin O, et al. The influence of dexmedetomidine on ischemic rat hippocampus [J]. Brain Res, 2008, 1218(7): 250-6.
- [24] Yoshitomi O, Cho S, Hara T, et al. Direct protective effects of dexmedetomidine against myocardial ischemia-reperfusion injury in anesthetized pigs [J]. Shock, 2012, 38(1): 92-7.
- [25] Sahin T, Begeç Z, Toprak Hİ, et al. The effects of dexmedetomidine on liver ischemia-reperfusion injury in rats [J]. J Surg Res, 2013, 183(1): 385-90.
- [26] Zhang XY, Liu ZM, Wen SH, et al. Dexmedetomidine administration before, but not after, ischemia attenuates intestinal injury induced by intestinal ischemia-reperfusion in rats [J]. Anesthesiology, 2012, 116(5): 1035-46.
- (编辑:吴锦雅)