

陈露,江小霞,朱玲玲.高原低压低氧环境下牙周炎与中枢神经系统炎症的相关性[J].中国比较医学杂志,2024,34(6):119-126.

Chen L, Jiang XX, Zhu LL. Correlation of periodontitis with central nervous system inflammation in hypobaric hypoxia environments at plateau [J]. Chin J Comp Med, 2024, 34(6): 119-126.

doi: 10.3969/j.issn.1671-7856.2024.06.016

高原低压低氧环境下牙周炎与中枢神经系统炎症的相关性

陈露^{1,2}, 江小霞^{2*}, 朱玲玲^{1,2*}

(1.南华大学衡阳医学院,军事科学院军事医学研究院研究生协作培养基地,湖南 衡阳 421001;

2.军事医学研究院,北京 100850)

【摘要】近年来,随着在高原和山区活动人群的增多,高海拔暴露变得越来越普遍,许多基础病患者受高原低压低氧环境影响,病程进一步加重甚至导致认知障碍的现象频发。牙周炎是一种常见的炎症性疾病,能诱发牙周局部炎症反应,甚至中枢神经系统炎症。在高海拔环境下,机体出现免疫力下降、组织缺氧等反应,这些反应会促进牙周炎的发生发展,甚至有可能增加牙周炎引发中枢神经系统炎症的风险。随着高原医学研究的不断深入,高原低压低氧环境下牙周炎与中枢神经系统炎症之间的关系引发越来越多的关注。本文就牙周炎与中枢神经系统炎症的研究进展进行综述,并对高原低压低氧环境下牙周炎与中枢神经系统炎症之间的相关性进行讨论。

【关键词】高原;低压低氧;牙周炎;中枢神经系统炎症

【中图分类号】R-33 【文献标识码】A 【文章编号】1671-7856(2024)06-0119-08

Correlation of periodontitis with central nervous system inflammation in hypobaric hypoxia environments at plateau

CHEN Lu^{1,2}, JIANG Xiaoxia^{2*}, ZHU Lingling^{1,2*}

(1. Graduate Collaborative Training Base of Academy of Military Medical Sciences, Academy of Military Sciences, Hengyang Medical School, University of South China, Hengyang 421001, China. 2. Academy of Military Medical Sciences, Beijing 100850)

【Abstract】With the increase in human activity in plateau and mountainous areas in recent years, high-altitude exposure has become increasingly common. Many patients with underlying diseases are affected by hypobaric hypoxia in plateau environments, which further aggravates disease processes and even leads to cognitive disorders. Periodontitis is a common inflammatory disease that induces periodontal local inflammatory responses and even causes central nervous system inflammation. At high altitudes, the body suffers from decreased immunity and tissue hypoxia, which can promote the occurrence and development of periodontitis and may even increase the risk of periodontitis-induced central nervous system inflammation. As plateau medical research advances, the relationship between periodontitis and central nervous system inflammation in hypobaric hypoxia environments at plateau is attracting more and more attention. This work reviews the

【基金项目】国家自然科学基金(82072104)。

【作者简介】陈露(1999—),女,硕士研究生,研究方向:高原医学研究。E-mail:chlu2971@163.com

【通信作者】朱玲玲(1966—),女,博士,研究员,研究方向:高原医学研究。E-mail:linglingzhuamms@126.com

江小霞(1976—),女,博士,副研究员,研究方向:情绪与认知调控研究。E-mail:smilovjiang@163.com * 共同通信作者

progress of research on periodontitis and central nervous system inflammation and discusses the correlation between periodontitis and central nervous system inflammation when exposed to hypobaric hypoxia in plateau environments.

【Keywords】 plateau; hypobaric hypoxia; periodontitis; central nervous system inflammation

Conflicts of Interest: The authors declare no conflict of interest.

随着社会的进步,迁移至高海拔地区就业和居住的人不断增多,低压低氧是高原地区的主要环境特征^[1],可以引发机体一系列不良反应^[2-4],甚至导致高原低压低氧脑损伤^[5-6]。而炎症在高原低压低氧脑损伤中发挥着重要作用^[7-8],系统性炎症和局部炎症的存在会增加高原低压低氧脑损伤的风险^[9-10]。牙周炎作为一种口腔局部炎症性疾病,以牙周袋壁炎症、牙槽骨吸收、牙齿松动为主要特征,高原低压低氧环境对牙周炎的病情会产生负面影响^[11],尽管已有研究发现牙周炎能够引发中枢神经系统炎症导致学习与记忆障碍^[12],但高原低压低氧环境下牙周炎与中枢神经系统炎症的讨论尚未见报道。因此,本文就高原低压低氧环境下牙周炎与中枢神经系统炎症的相关性进行讨论,或可为高原低压低氧环境下牙周炎和中枢神经系统炎症的预防提供思路。

1 牙周炎的现状

牙周炎是由牙周病原微生物引发的牙周组织炎症性疾病,初期牙龈轻微出血,随着时间的推移,牙菌斑生长,牙结石积聚,牙周炎症加重,最终导致牙齿脱落^[13]。除了微生物因素外,环境、宿主炎症和遗传等因素也推动着牙周炎的发生和发展^[14],使牙周炎在人群中广泛流行。我国第四次全国口腔疾病流行病学调查显示,牙周炎影响了一半以上中国成年人的口腔健康^[15-16]。在全球范围内,约 7.43 亿人的口腔健康受牙周炎影响^[17],且患病率呈上升趋势^[18],牙周炎已成为全球口腔健康问题。不仅如此,研究发现牙周炎还是中枢神经系统炎症的重要危险因素之一^[12]。

2 牙周炎对中枢神经系统炎症的影响

长期以来,中枢神经系统炎症是中枢神经系统疾病的重要病理机制^[19],牙周炎作为一种局部炎症与中枢神经系统炎症密切相关^[20-21]。而牙周炎引发中枢神经系统炎症的主要病因一方面是牙周病原微生物及其产物^[20],包括牙龈卟啉单胞菌(*Porphyromonas gingivalis*, Pg)^[22]、齿垢密螺旋体(*Treponema denticola*, Td)^[23]、具核梭杆菌

(*Fusobacterium nucleatum*, Fn)^[24]等,这些病原微生物进入脑内能引发免疫反应,还能产生多种毒力因子激活脑内免疫细胞,如脂多糖(lipopolysaccharide, LPS)^[25-26]、牙龈蛋白酶^[27-28]、荚膜^[29]、外膜囊泡(outer membrane vesicle, OMV)^[30]、新型鞘脂^[31]。另一方面,牙周炎症状态时释放的炎症因子一旦进入大脑,不仅会增加炎症因子的数量,也会刺激神经胶质细胞产生额外的炎症因子,引发脑内炎症反应^[32]。这些致炎物质从牙周组织出发,通过多种途径进入脑内(图 1),主要分为以下 4 种途径:

(1) 血液循环途径:研究显示,牙龈卟啉单胞菌通过牙周袋渗漏进入血液循环,一旦进入脑血管,可能会通过多种方式诱导血脑屏障(blood brain barrier, BBB)的破坏,使 Pg 经血脑屏障进入脑内^[32-33]。牙周炎症部位还能释放炎症因子进入血液循环,炎症因子穿过血脑屏障进入脑内^[34]。不仅如此,脑室周围器官(circumventricular organs, CVO)缺乏连续的血脑屏障,当牙周病原微生物和炎症因子进入血液后可能经此位置进入大脑^[35]。另外,大脑表面蛛网膜与软脑膜间的蛛网膜下腔含有丰富的血管^[36],Pg-LPS 经血液循环到达脑血管后能使外周巨噬细胞产生炎症因子,软脑膜细胞能够将外周巨噬细胞炎症信号转导到脑内^[37],炎症因子还能与脑血管内皮细胞(endothelial cells, EC)上的炎症因子受体结合,激活血管周围巨噬细胞与小胶质细胞间的通讯,使小胶质细胞分泌炎症因子,导致神经炎症^[38]。

(2) 神经旁路途径:齿垢密螺旋体是牙周炎的病原微生物之一^[39],能通过三叉神经进入大脑到达三叉神经中脑核和海马体^[40-41],也能沿着嗅神经进入大脑^[35]。牙龈卟啉单胞菌细胞外囊泡(extracellular vesicles, EVs)能通过三叉神经进入大脑,诱发神经炎症^[42]。此外,外周炎症因子还能刺激周围神经传入纤维,激活迷走神经将信号传入大脑,导致脑内炎症因子水平升高^[43]。

(3) 口腔微生物-肠-脑轴途径:研究发现,肠道微生物通过肠-脑轴影响其宿主的大脑功能,涉及神经、内分泌和免疫途径^[44],而口腔菌群变化会引起肠道菌群的改变^[45]。因此,口腔微生物通过影响

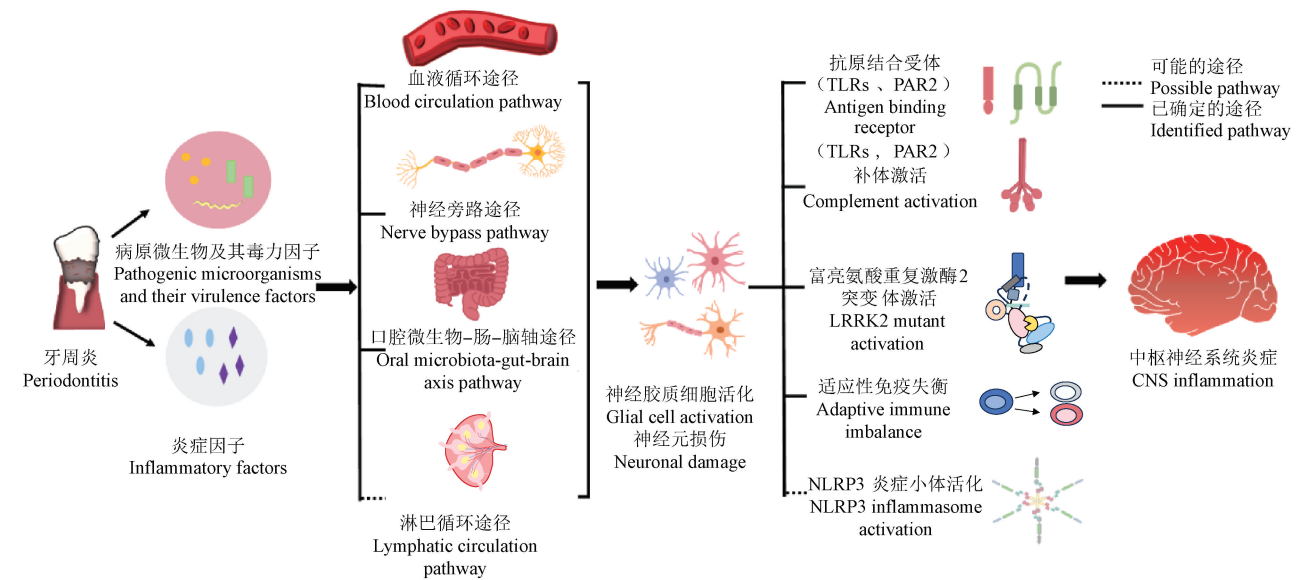


图 1 牙周炎影响中枢神经系统炎症的途径

Figure 1 Pathways of periodontitis affecting CNS inflammation

肠-脑轴从而影响中枢神经系统炎症发生是有可能的。在小鼠模型中,牙周炎相关唾液微生物群的持续灌胃损害认知功能,导致脑内神经炎症^[46]。Xue 等^[47]发现小鼠慢性牙周炎诱导口腔微生物-肠-脑轴紊乱导致中枢神经系统炎症与认知障碍也为此途径提供了直接证据。

(4) 淋巴循环途径:淋巴管中含有大量抗原呈递细胞,能识别抗原并呈递至区域淋巴结进行处理^[48],其中第三和第四脑室与颈部中深淋巴结相连^[49-50],口腔区域淋巴也与颈部中深淋巴结相连^[32],虽然目前没有证据表明它是将口腔细菌传播到大脑的通路,但是是一种可能的途径。

以上研究表明,牙周病原微生物及其产物和炎症因子能通过多种方式侵入脑内,引发中枢神经系统炎症。而作为脑内的主要免疫细胞,小胶质细胞和星形胶质细胞在大脑中有调节神经炎症的作用,是脑内炎症因子的重要来源^[51]。在牙周炎与中枢神经系统炎症的联系中,小胶质细胞和星形胶质细胞的活化占据重要地位。根据文献报道,牙周病原微生物及其产物和炎症因子使小胶质细胞和星形胶质细胞活化,引发神经元损伤,导致脑内神经炎症的具体机制包括以下 5 点:

(1) 抗原结合受体:研究发现,Pg-LPS 能通过结合细胞膜上的 Toll 样受体,激活 NF- κ B/STAT3 信号通路,上调炎症因子表达,活化小胶质细胞和星形胶质细胞^[25,52]。此外,Pg 产生的牙龈蛋白酶能与蛋白酶激活受体 2 (proteinase activated receptor-2,

PAR2) 结合,激活 PI3K/AKT 信号通路和 ERK 信号通路,使小胶质细胞释放 IL-6 和 TNF- α ^[28]。

(2) 补体激活:在阿尔茨海默病 (Alzheimer's disease, AD) 小鼠中, Pg 能够穿过血脑屏障进入脑内,激活小胶质细胞产生炎症因子,并且还能够诱导补体成分 1q (complement component 1q, C1q) 过表达,使小胶质细胞进一步活化,加剧中枢神经系统炎症^[53]。

(3) 富亮氨酸重复激酶 2 (leucine-rich repeat kinase 2, LRRK2) 突变体激活:富亮氨酸重复激酶 2 突变体是帕金森病发病机制中最常见的遗传因素^[54]。研究发现, Pg 能激活 LRRK2 突变体,使小胶质细胞活化,诱导的神经变性具有 LRRK2 突变依赖性^[55]。

(4) 适应性免疫失衡:适应性免疫的主要参与者是 CD4⁺T 细胞,通过分化为辅助性 T 细胞 (helper T cell, Th) 和调节性 T 细胞 (regulatory T cell, Treg), 可以调控脑内神经炎症^[56]。研究发现,在 Pg-LPS 诱导的牙周炎小鼠中,Th17 相关促炎细胞因子在血液和大脑中的表达增加,而 Treg 相关抗炎细胞因子的表达降低,神经元凋亡加剧^[57]。

(5) NLRP3 炎症小体活化:NOD 样受体热蛋白结构域相关蛋白 3 (NOD-like receptor thermal protein domain associated protein 3, NLRP3) 炎症小体是先天免疫系统的关键部分,在多种炎症性疾病中起重要作用^[58]。Gong 等^[30]研究发现,口服饲喂小鼠牙龈卟啉单胞菌外膜囊泡后,可在海马体和皮层中检测

到外膜囊泡,星形胶质细胞和小胶质细胞活化,IL-1 β 水平增加,海马体中 NLRP3 炎症小体相关蛋白的表达上调,表明外膜囊泡能激活脑内免疫细胞进而引发中枢神经系统炎症,NLRP3 炎症小体激活是一种可能的机制。这些研究进一步证明牙周炎是中枢神经系统炎症的重要危险因素之一。

3 高原低压低氧环境下牙周炎与中枢神经系统炎症的关系

随着高原医学研究的深入,科学家们开始探索高原低压低氧环境下牙周炎与中枢神经系统炎症之间的可能关联。目前关于这方面的研究,杜灿等^[59]讨论了高原慢性低氧环境慢性牙周炎与阿尔茨海默病的相关性,提出了相关见解,但高原低压低氧环境下牙周炎与中枢神经系统炎症的研究目前仍未见报道。因此,接下来围绕以下两个部分对高原低压低氧环境下牙周炎与中枢神经系统炎症相关关系进行讨论,为进一步的实验研究提供思路。

3.1 高原低压低氧环境对牙周炎的影响

近年来,高原低压低氧环境对牙周炎的影响引发关注。根据我国口腔流行病学调查统计,平原地区生活人群牙结石、牙龈出血检出率分别为 53.5%、18.8%,而西藏地区的高原移民检出率为 68.6%、30.4%^[60]。不仅如此,在一项针对中国西部地区青少年牙龈状况与海拔的相关性分析中发现,64.09%和 77.15%的青少年分别患有牙龈出血和牙结石,高原地区青少年患病率显著高于平原地区青少年^[61]。以上研究均表明,高原环境下牙周炎的发病率明显高于平原地区。此外,一项对空军部队的口腔健康状况调查分析也显示,驻高原空军官兵牙周炎患病率显著高于平原空军官兵,且随高原驻军时间延长,牙周炎逐渐加重^[62],这提示高原环境与牙周炎发病率和病情发展相关。

首先,在人群研究中发现,高原牙周炎患者血清中内脂素(visfatin, VF)水平升高,促炎作用增强,导致牙周组织损伤加重^[63]。此外,在牙周炎兔模型中,低压低氧后血清和牙龈组织中细胞间黏附分子-1(soluble intercellular adhesion molecule-1, sICAM-1)、可溶性血管黏附分子-1(soluble vascular cell adhesion molecule-1, sVCAM-1)显著上调^[64],提示血管通透性增加;超氧化物歧化酶(superoxide dismutase, SOD)活力降低,超氧自由基(superoxide radical, SR)增多,炎症加剧^[65]。而在牙周炎大鼠模型中,低压低氧后牙周组织炎症因子水平显著上

升^[66],龈沟液中 C-反应蛋白(C-reactive protein, CRP)水平增加^[67],血清中基质金属蛋白酶-2(matrix metalloproteinase-2, MMP-2)和基质金属蛋白酶-3(matrix metalloproteinase-3, MMP-3)表达上调^[68-69],增强牙周组织细胞外基质的降解,使炎症进一步发展。以上人群和实验动物模型的研究结果均表明低压低氧能够加速牙周炎的发生发展,而高原低压低氧加重牙周炎症的主要原因可能涉及以下 4 个方面:

(1) 低压低氧使牙周病原微生物增多

牙周病原微生物是导致牙周炎的主要因素,黄镜静等^[70]研究发现牙周炎兔模型低压低氧后龈下菌斑中牙龈卟啉单胞菌水平相比其余各组显著升高。李森等^[71]采用高通量测序检测菌群,发现高原牙周炎人群口腔菌群中齿垢密螺旋体等专性厌氧菌增多,表明低压低氧有利于牙周病原微生物生长。而牙周病原微生物能产生多种毒力成分^[72],使口腔微生物生态失调,诱发牙周组织炎症。

(2) 低压低氧加重牙周组织局部缺血缺氧

人在高原低压低氧条件下血氧饱和度(oxygen saturation, SaO₂)远低于正常水平^[73],为保证机体供氧,人体产生更多红细胞和血红蛋白以捕捉氧气分子,从而引发小血管破裂,使局部组织缺血缺氧,引发无氧代谢,细菌大量繁殖^[74]。此外,牙周组织缺氧会抑制人牙周膜细胞(human periodontal ligament cells, hPDLs)的增殖和迁移能力^[75]。同时,缺氧也会引发自由基累积,炎症因子大量释放^[65-66],使牙周炎症加重。

(3) 低压低氧破坏牙周组织防御功能

正常状态下,口腔中的中性粒细胞能够发挥杀灭病原微生物的作用^[76],而在高原低压低氧条件下牙周炎患者产生的中性粒细胞减少,识别和清除病原微生物的效果减弱,加速牙周组织感染发炎^[11]。

(4) 高原环境其他特点加重牙周炎症

高原地区与平原相比风力、昼夜温差和日照强度明显增强,当上升到高原后唾液分泌无法达到正常水平,导致口腔清洁能力减弱,牙周微生物加快繁殖。此外,高原地区饮食以肉奶制品为主,蔬菜水果食用频率较低,缺乏人体所需的维生素和微量元素,易导致口腔疾病的发生。同时,高原水中丰富的矿物质易沉积在牙齿表面,导致牙结石的形成,加重牙周炎^[77]。

以上研究表明高原低压低氧环境会加重牙周

局部炎症,而牙周炎症引发中枢神经系统炎症的研究已有报道,但在高原特殊环境下牙周炎与中枢神经系统炎症之间的关系更为复杂。

3.2 高原低压低氧环境下牙周炎与中枢神经系统炎症的相关性

近年来,高原低压低氧环境下牙周炎与中枢神经系统炎症的关系引发关注,赵婷婷^[78]研究发现牙周炎能够导致藏族中老年人认知功能明显下降,提示慢性牙周炎有可能是高原地区认知障碍的危险因素。此外,2018 年报道了一例与海拔升高相关的牙周感染导致脑脓肿的病例,表明海拔升高后低压环境对血流动力学的长期影响会导致牙周菌群分流,从而导致颅内神经炎症的发生^[79]。以上研究表明高原低压低氧环境下牙周炎与中枢神经系统炎症相关,并且高原低压低氧环境可能对牙周炎引发中枢神经系统炎症具有促进作用,虽未阐明具体机制,但为高原低压低氧环境下牙周炎与中枢神经系统炎症的研究做出了初步探索。

4 展望

目前,高原低压低氧环境下牙周炎与中枢神经系统炎症之间的研究尚有待进一步深入。虽然已有研究表明高原低压低氧能够加重牙周炎,且牙周炎与中枢神经系统炎症之间存在关联,但仅限于对平原地区牙周炎与中枢神经系统炎症的研究较多,高原地区不仅对于牙周炎的研究较少,对于牙周炎与中枢神经系统炎症的研究更少,特别是在高原低压低氧环境下的特殊变化及其变化规律,与高海拔暴露的时间的关联性等都有待深入探究。此外,牙周炎进程包含不同阶段,不同阶段牙周炎在低压低氧环境下对中枢神经系统炎症的影响是否不同,牙周炎处于何种阶段在高原低压低氧环境下会引发中枢神经系统炎症,这些问题尚待进一步的研究。另外,中枢神经系统炎症是引发神经退行性疾病的重要原因,并且在高原环境下与高原脑水肿等高原病也有关联。因此,高原环境下牙周炎与神经退行性疾病和高原病的关系均是需要进一步探讨的问题。当牙周炎与高原低压低氧两种风险因素都存在时,发生中枢神经系统炎症的风险是否增加?是否会引发神经退行性疾病和高原病?其中的具体机制是什么?对这些问题的研究将阐明高原低压低氧环境下牙周炎对中枢神经系统炎症以及相关疾病发生和发展的影响,从而为高原低压低氧环境

下中枢神经系统炎症的预防与治疗提供思路,为到高海拔地区的人群的健康提供有价值的建议。

参考文献:

- [1] 范明, 朱玲玲. 低氧研究受到关注 [J]. 生理学报, 2019, 71(5): 806-808.
FAN M, ZHU L L. Hypoxia research receives attention [J]. Acta Physiol Sin, 2019, 71(5): 806-808.
- [2] 向雪梅, 郭鑫, 曾佳容, 等. 高原肺水肿动物模型研究进展 [J]. 中国实验动物学报, 2022, 30(2): 291-298.
XIANG X M, GUO X, ZENG J R, et al. Evolution of animal models of high altitude pulmonary edema [J]. Acta Lab Anim Sci Sin, 2022, 30(2): 291-298.
- [3] 付鹏宇, 胡扬, 李燕春, 等. 低氧暴露所致大鼠骨骼肌萎缩的蛋白转化调节机制 [J]. 中国实验动物学报, 2019, 27(4): 423-432.
FU P Y, HU Y, LI Y C, et al. Protein turnover regulation mechanism of rat skeletal muscle atrophy induced by hypoxia [J]. Acta Lab Anim Sci Sin, 2019, 27(4): 423-432.
- [4] LI F, QIAO Z, DUAN Q, et al. Adaptation of mammals to hypoxia [J]. Animal Model Exp Med, 2021, 4(4): 311-318.
- [5] 张梦雅, 赵名, 江亚群, 等. 慢性高原低氧暴露对小鼠神经行为学的影响 [J]. 中国比较医学杂志, 2023, 33(4): 28-35.
ZHANG M Y, ZHAO M, JIANG Y Q, et al. Effects of chronic high-altitude hypoxia exposure on neurobehavior in mice [J]. Chin J Comp Med, 2023, 33(4): 28-35.
- [6] 刘鹏飞, 胡艳婷, 姜静雯, 等. 通心络胶囊对大鼠低压低氧暴露后炎症反应和脑组织水肿及认知功能的影响 [J]. 中国比较医学杂志, 2021, 31(6): 69-76.
LIU P F, HU Y T, JIANG J W, et al. Effects of tongxinluo capsule on hippocampal inflammation, brain edema, and cognitive function of rats exposed to acute hypobaric hypoxia [J]. Chin J Comp Med, 2021, 31(6): 69-76.
- [7] PHAM K, PARIKH K, HEINRICH E C. Hypoxia and inflammation: insights from high-altitude physiology [J]. Front Physiol, 2021, 12: 676782.
- [8] 史子毕. IL-6 与高原病的研究进展 [J]. 中国高原医学与生物学杂志, 2021, 42(3): 190-194.
SHI Z B. Research progress of IL-6 and altitude sickness [J]. Chin High Alt Med Biol, 2021, 42(3): 190-194.
- [9] ZHOU Y, HUANG X, ZHAO T, et al. Hypoxia augments LPS-induced inflammation and triggers high altitude cerebral edema in mice [J]. Brain Behav Immun, 2017, 64: 266-275.
- [10] HAN Y, DING L, CHENG X, et al. Hypoxia augments cerebral inflammation in a dextran sulfate sodium-induced colitis mouse model [J]. Front Cell Neurosci, 2020, 14: 611764.
- [11] 李婧. 高原地理环境与牙周炎的关系研究进展 [J]. 全科口腔医学杂志(电子版), 2020, 7(1): 26, 36.
LI J. Research progress on the relationship between plateau geographical environment and periodontitis [J]. Electron J Gen

- Stomatol, 2020, 7(1): 26, 36.
- [12] HU Y, LI H, ZHANG J, et al. Periodontitis induced by *P. gingivalis*-LPS is associated with neuroinflammation and learning and memory impairment in sprague-dawley rats [J]. Front Neurosci, 2020, 14: 658.
 - [13] JOHNSON A, HE J L, KONG F, et al. Surfactin-loaded K-carrageenan oligosaccharides entangled cellulose nanofibers as a versatile vehicle against periodontal pathogens [J]. Int J Nanomedicine, 2020, 15: 4021–4047.
 - [14] TELES F, COLLMAN R G, MOMINKHAN D, et al. Viruses, periodontitis, and comorbidities [J]. Periodontol 2000, 2022, 89(1): 190–206.
 - [15] SUN H Y, JIANG H, DU M Q, et al. The prevalence and associated factors of periodontal disease among 35 to 44-year-old Chinese adults in the 4th national oral health survey [J]. Chin J Dent Res, 2018, 21(4): 241–247.
 - [16] SUN H, DU M, TAI B, et al. Prevalence and associated factors of periodontal conditions among 55- to 74-year-old adults in China: results from the 4th National Oral Health Survey [J]. Clin Oral Investig, 2020, 24(12): 4403–4412.
 - [17] YOST S, DURAN-PINEDO A E, TELES R, et al. Functional signatures of oral dysbiosis during periodontitis progression revealed by microbial metatranscriptome analysis [J]. Genome Med, 2015, 7(1): 27.
 - [18] YAN Y, ZHAN Y, WANG X, et al. Clinical evaluation of ultrasonic subgingival debridement versus ultrasonic subgingival scaling combined with manual root planing in the treatment of periodontitis; study protocol for a randomized controlled trial [J]. Trials, 2020, 21(1): 113.
 - [19] MITHAIWALA M N, SANTANA-COELHO D, PORTER G A, et al. Neuroinflammation and the kynurenine pathway in CNS disease: molecular mechanisms and therapeutic implications [J]. Cells, 2021, 10(6): 1548.
 - [20] 刘珂珂, 胡韶光, 吕代雨, 等. 结扎诱导牙周炎大鼠脑内炎症性及 β 淀粉样蛋白变化的实验研究 [J]. 安徽医科大学学报, 2022, 57(7): 1048–1053.
 - LIU K K, HU S G, LYU D Y, et al. Changes of inflammatory and β -amyloid in the brain of rats with periodontitis induced by ligation [J]. Acta Univ Med Anhui, 2022, 57(7): 1048–1053.
 - [21] WANG R P, HUANG J, CHAN K W Y, et al. IL-1 β and TNF- α play an important role in modulating the risk of periodontitis and Alzheimer's disease [J]. J Neuroinflammation, 2023, 20(1): 71.
 - [22] TANG Z, LIANG D, CHENG M, et al. Effects of *Porphyromonas gingivalis* and its underlying mechanisms on Alzheimer-like tau hyperphosphorylation in Sprague-Dawley rats [J]. J Mol Neurosci, 2021, 71(1): 89–100.
 - [23] WU L, SU X, TANG Z, et al. *Treponema denticola* induces neuronal apoptosis by promoting amyloid- β accumulation in mice [J]. Pathogens, 2022, 11(10): 1150.
 - [24] YAN C, DIAO Q, ZHAO Y, et al. *Fusobacterium nucleatum* infection-induced neurodegeneration and abnormal gut microbiota composition in Alzheimer's disease-like rats [J]. Front Neurosci, 2022, 16: 884543.
 - [25] ZHANG J, YU C, ZHANG X, et al. *Porphyromonas gingivalis* lipopolysaccharide induces cognitive dysfunction, mediated by neuronal inflammation via activation of the TLR4 signaling pathway in C57BL/6 mice [J]. J Neuroinflammation, 2018, 15(1): 37.
 - [26] WU Z, NI J, LIU Y, et al. Cathepsin B plays a critical role in inducing Alzheimer's disease-like phenotypes following chronic systemic exposure to lipopolysaccharide from *Porphyromonas gingivalis* in mice [J]. Brain Behav Immun, 2017, 65: 350–361.
 - [27] HADITSCH U, ROTH T, RODRIGUEZ L, et al. Alzheimer's disease-like neurodegeneration in *Porphyromonas gingivalis* infected neurons with persistent expression of active gingipains [J]. J Alzheimers Dis, 2020, 75(4): 1361–1376.
 - [28] LIU Y, WU Z, NAKANISHI Y, et al. Infection of microglia with *Porphyromonas gingivalis* promotes cell migration and an inflammatory response through the gingipain-mediated activation of protease-activated receptor-2 in mice [J]. Sci Rep, 2017, 7(1): 11759.
 - [29] DÍAZ-ZÚÑIGA J, MORE J, MELGAR-RODRÍGUEZ S, et al. Alzheimer's disease-like pathology triggered by *Porphyromonas gingivalis* in wild type rats is serotype dependent [J]. Front Immunol, 2020, 11: 588036.
 - [30] GONG T, CHEN Q, MAO H, et al. Outer membrane vesicles of *Porphyromonas gingivalis* trigger NLRP3 inflammasome and induce neuroinflammation, tau phosphorylation, and memory dysfunction in mice [J]. Front Cell Infect Microbiol, 2022, 12: 925435.
 - [31] YAMADA C, AKKAOUI J, HO A, et al. Potential role of phosphoglycerol dihydroceramide produced by periodontal pathogen *Porphyromonas gingivalis* in the pathogenesis of Alzheimer's disease [J]. Front Immunol, 2020, 11: 591571.
 - [32] SANSORES-ESPAÑA D, CARRILLO-AVILA A, MELGAR-RODRIGUEZ S et al. Periodontitis and alzheimer's disease [J]. Med Oral Patol Oral Cir Bucal, 2021, 26(1): e43–e48.
 - [33] NONAKA S, KADOWAKI T, NAKANISHI H. Secreted gingipains from *Porphyromonas gingivalis* increase permeability in human cerebral microvascular endothelial cells through intracellular degradation of tight junction proteins [J]. Neurochem Int, 2022, 154: 105282.
 - [34] FURUTAMA D, MATSUDA S, YAMAWAKI Y, et al. IL-6 induced by periodontal inflammation causes neuroinflammation and disrupts the blood-brain barrier [J]. Brain Sci, 2020, 10(10): 679.
 - [35] RANJAN R, ABHINAY A, MISHRA M. Can oral microbial infections be a risk factor for neurodegeneration? A review of the literature [J]. Neurol India, 2018, 66(2): 344–351.
 - [36] WELLER R O, SHARP M M, CHRISTODOULIDES M, et al. The meninges as barriers and facilitators for the movement of

- fluid, cells and pathogens related to the rodent and human CNS [J]. Acta Neuropathol, 2018, 135(3): 363-385.
- [37] LIU Y, WU Z, ZHANG X, et al. Leptomeningeal cells transduce peripheral macrophages inflammatory signal to microglia in reponse to *Porphyromonas gingivalis* LPS [J]. Mediators Inflamm, 2013, 2013: 407562.
- [38] D'MELLO C, SWAIN M G. Immune-to-brain communication pathways in inflammation-associated sickness and depression [J]. Curr Top Behav Neurosci, 2017, 31: 73-94.
- [39] LI R, WANG J, XIONG W, et al. The oral-brain axis: can periodontal pathogens trigger the onset and progression of Alzheimer's disease? [J]. Front Microbiol, 2024, 15: 1358179.
- [40] PISANI F, PISANI V, ARCANGELI F, et al. The mechanistic pathways of periodontal pathogens entering the brain: the potential role of *Treponema denticola* in tracing Alzheimer's disease pathology [J]. Int J Environ Res Public Health, 2022, 19(15): 9386.
- [41] RIVIERE G R, RIVIERE K H, SMITH K S. Molecular and immunological evidence of oral *Treponema* in the human brain and their association with Alzheimer's disease [J]. Oral Microbiol Immunol, 2002, 17(2): 113-118.
- [42] HA J Y, SEOK J, KIM S J, et al. Periodontitis promotes bacterial extracellular vesicle-induced neuroinflammation in the brain and trigeminal ganglion [J]. PLoS Pathog, 2023, 19(10): e1011743.
- [43] 边梦瑶, 陈莉丽, 雷利红. 慢性牙周炎与帕金森病相关性的研究进展 [J]. 浙江大学学报(医学版), 2022, 51(1): 108-114.
BIAN M Y, CHEN L L, LEI L H. Research progress on the relationship between chronic periodontitis and Parkinson's disease [J]. J Zhejiang Univ Med Sci, 2022, 51(1): 108-114.
- [44] SUDA K, MATSUDA K. How microbes affect depression: underlying mechanisms via the gut-brain axis and the modulating role of probiotics [J]. Int J Mol Sci, 2022, 23(3): 1172.
- [45] HAN Y, WANG B, GAO H, et al. Insight into the relationship between oral microbiota and the inflammatory bowel disease [J]. Microorganisms, 2022, 10(9): 1868.
- [46] LU J, ZHANG S, HUANG Y, et al. Periodontitis-related salivary microbiota aggravates Alzheimer's disease via gut-brain axis crosstalk [J]. Gut Microbes, 2022, 14(1): 2126272.
- [47] XUE L, ZOU X, YANG X Q, et al. Chronic periodontitis induces microbiota-gut-brain axis disorders and cognitive impairment in mice [J]. Exp Neurol, 2020, 326: 113176.
- [48] MAGOLD A I, SWARTZ M A. Pathogenic exploitation of lymphatic vessels [J]. Cells, 2022, 11(6): 979.
- [49] LOUVEAU A. Cerebral lymphatic drainage: implication in the brain immune privilege [J]. Med Sci, 2015, 31(11): 953-956.
- [50] LOUVEAU A, SMIRNOV I, KEYES T J, et al. Structural and functional features of central nervous system lymphatic vessels [J]. Nature, 2015, 523(7560): 337-341.
- [51] TANAKA T, KAI S, MATSUYAMA T, et al. General anesthetics inhibit LPS-induced IL-1 β expression in glial cells [J]. PLoS One, 2013, 8(12): e82930.
- [52] QIU C, YUAN Z, HE Z, et al. Lipopolysaccharide preparation derived from *Porphyromonas gingivalis* induces a weaker Immuno-inflammatory response in BV-2 microglial cells than *Escherichia coli* by differentially activating TLR2/4-mediated NF- κ B/STAT3 signaling pathways [J]. Front Cell Infect Microbiol, 2021, 11: 606986.
- [53] HAO X, LI Z, LI W, et al. Periodontal infection aggravates C1q-mediated microglial activation and synapse pruning in Alzheimer's mice [J]. Front Immunol, 2022, 13: 816640.
- [54] CHANG E E S, HO P W, LIU H F, et al. LRRK2 mutant knock-in mouse models: therapeutic relevance in Parkinson's disease [J]. Transl Neurodegener, 2022, 11(1): 10.
- [55] FENG Y K, WU Q L, PENG Y W, et al. Oral *P. gingivalis* impairs gut permeability and mediates immune responses associated with neurodegeneration in LRRK2 R1441G mice [J]. J Neuroinflammation, 2020, 17(1): 347.
- [56] CAMPOS-ACUÑA J, ELGUETA D, PACHECO R. T-cell-driven inflammation as a mediator of the gut-brain axis involved in Parkinson's disease [J]. Front Immunol, 2019, 10: 239.
- [57] ZHANG X, ZHANG X, QIU C, et al. The imbalance of Th17/Treg via STAT3 activation modulates cognitive impairment in *P. gingivalis* LPS-induced periodontitis mice [J]. J Leukoc Biol, 2021, 110(3): 511-524.
- [58] FU J, WU H. Structural mechanisms of NLRP3 inflammasome assembly and activation [J]. Annu Rev Immunol, 2023, 41: 301-316.
- [59] 杜灿, 武洲, 朱爱琴. 高原慢性低氧环境慢性牙周炎与阿尔茨海默病的相关性 [J]. 中华老年医学杂志, 2015, 34(4): 452-454.
DU C, WU Z, ZHU A Q. The correlation of Alzheimer disease and chronic periodontitis in chronic hypoxia environment at plateau [J]. Chin J Geriatr, 2015, 34(4): 452-454.
- [60] 肖娴, 张纲, 李焰, 等. 驻高原和平原官兵口腔健康状况调查 [J]. 解放军预防医学杂志, 2010, 28(1): 40-41.
XIAO X, ZHANG G, LI Y, et al. Investigation on oral health status of officers and men stationed in plateau and plain [J]. J Prev Med Chin People's Liberation Army, 2010, 28(1): 40-41.
- [61] WU Z, ZHANG R, CHENG L, et al. The correlation of altitude with gingival status among adolescents in Western China: a cross-sectional study [J]. Environ Geochem Health, 2021, 43(8): 3151-3167.
- [62] 余建祯, 冯帆, 周兴田, 等. 驻高原与驻平原空军官兵口腔健康现状调查 [J]. 西南军医, 2016, 18(2): 101-103.
SHE J Z, FENG F, ZHOU X T, et al. Oral health status of air force personnel on plateau and on plain [J]. J Mil Surg Southwest China, 2016, 18(2): 101-103.
- [63] 李燕, 张羽涛, 刘倩雯, 等. 高原地区牙周炎患者血清内脂

- 素和瘦素水平的检测分析 [J]. 口腔医学, 2022, 42(1): 58-61.
- LI Y, ZHANG Y T, LIU Q W, et al. Serum visfatin and leptin levels and their associations with periodontal indexes in patients with periodontitis in plateau section [J]. Stomatology, 2022, 42(1): 58-61.
- [64] 武曦, 黄镜静, 张纲, 等. 模拟高原低氧兔牙周炎模型血清和牙龈组织中 sICAM-1 及 sVCAM-1 的含量 [J]. 实用口腔医学杂志, 2012, 28(2): 178-181.
- WU X, HUANG J J, ZHANG G, et al. sICAM-1 and sVCAM-1 levels in blood serum and gingival tissues in rabbits with periodontitis in analogic high altitude anoxia environment [J]. J Pract Stomatol, 2012, 28(2): 178-181.
- [65] 武曦, 黄镜静, 张纲, 等. 高原低氧环境对兔牙周炎模型血清和牙龈组织中超氧化物歧化酶活力的影响 [J]. 华西口腔医学杂志, 2012, 30(3): 247-250.
- WU X, HUANG J J, ZHANG G, et al. Effects of the activity of superoxide dismutase in blood serum and gingival tissues of rabbit in periodontitis model after hypoxia exposure at high altitude [J]. West China J Stomatol, 2012, 30(3): 247-250.
- [66] 张丹丹, 许晓虎. 牙周炎大鼠 MMP-2、BGP 及炎性细胞因子的表达特点及低氧对于相关因子的影响 [J]. 医学研究杂志, 2018, 47(6): 176-179.
- ZHANG D D, XU X H. Effect of hypoxia on periodontitis and the effect of MMP-2, BGP and inflammatory cytokines [J]. J Med Res, 2018, 47(6): 176-179.
- [67] 周霞, 刘鲁川, 孔耀, 等. 模拟高原条件下牙周炎动物模型龈沟液中 C-反应蛋白的变化 [J]. 第三军医大学学报, 2010, 32(2): 107-110.
- ZHOU X, LIU L C, KONG Y, et al. Gingival crevicular fluid C-reactive protein level in periodontitis rats living in simulated plateau-condition [J]. J Army Med Univ, 2010, 32(2): 107-110.
- [68] 孔耀, 刘鲁川, 周霞, 等. 模拟高原缺氧对慢性牙周炎大鼠血清中 MMP-2 表达的影响 [J]. 第三军医大学学报, 2010, 32(4): 324-326.
- KONG Y, LIU L C, ZHOU X, et al. Effect of simulated high altitude hypoxia on matrix metalloproteinase-2 expression in serum of rats with chronic periodontitis [J]. J Army Med Univ, 2010, 32(4): 324-326.
- [69] 孔耀, 刘鲁川, 周霞, 等. 模拟高原缺氧对慢性牙周炎大鼠血清中 MMP-3 含量变化的影响 [J]. 牙体牙髓牙周病学杂志, 2009, 19(12): 677-680, 725.
- KONG Y, LIU L C, ZHOU X, et al. Effect of the level of MMP-3 in serum of chronic periodontitis model in rat exposed to hypoxia simulating high altitude [J]. Chin J Conservative Dent, 2009, 19(12): 677-680, 725.
- [70] 黄镜静, 武曦, 张纲, 等. 模拟高原低氧环境对牙龈卟啉单胞菌的影响 [J]. 解放军医学杂志, 2012, 37(4): 375-378.
- HUANG J J, WU X, ZHANG G, et al. Effect of simulated high-altitude hypoxia on *Porphyromonas gingivalis* [J]. Med J Chin People's Liberation Army, 2012, 37(4): 375-378.
- [71] 李森, 张文瑾, 王思凡, 等. 甘肃省高原环境中人群牙周菌群研究 [J]. 兰州大学学报(医学版), 2022, 48(10): 52-56.
- LI M, ZHANG W J, WANG S F, et al. Analysis of periodontal flora in plateau environment of Gansu Province [J]. J Lanzhou Univ Med Sci, 2022, 48(10): 52-56.
- [72] HOW K Y, SONG K P, CHAN K G. *Porphyromonas gingivalis*: an overview of periodontopathic pathogen below the gum line [J]. Front Microbiol, 2016, 7: 53.
- [73] LI C, LI X, LIU J, et al. Investigation of the differences between the Tibetan and Han populations in the hemoglobin-oxygen affinity of red blood cells and in the adaptation to high-altitude environments [J]. Hematology, 2018, 23(5): 309-313.
- [74] 武曦, 张纲, 谭颖微. 高原低氧环境下牙周炎的发病机制 [J]. 现代生物医学进展, 2011, 11(13): 2587-2589, 2593.
- WU X, ZHANG G, TAN Y H. Pathogenesis of periodontitis in Anoxia and high altitude environment [J]. Prog Mod Biomed, 2011, 11(13): 2587-2589, 2593.
- [75] 萧智利. 低氧诱导 HIF1 α 抑制 hPDLcs 生物学活性及牙周炎组织中差异表达 LncRNA 的筛选 [D]. 重庆: 重庆医科大学, 2019.
- XIAO Z L. Hypoxia-induced HIF-1 α inhibits the biological activity of hPDLcs and screening the differentially expressed LncRNA in periodontitis tissues [D]. Chongqing: Chongqing Medical University, 2019.
- [76] BASSANI B, CUCCHIARA M, BUTERA A, et al. Neutrophils' contribution to periodontitis and periodontitis-associated cardiovascular diseases [J]. Int J Mol Sci, 2023, 24(20): 15370.
- [77] 何雪娇, 蒋俊强, 罗勇军. 高原地理环境与牙周炎的关系研究进展 [J]. 国外医学(医学地理分册), 2015, 36(2): 102-104.
- HE X J, JIANG J Q, LUO Y J. Research progress in the relationship between plateau geographical environment and periodontitis [J]. Foreign Med Sci Sect Medgeogr, 2015, 36(2): 102-104.
- [78] 赵婷婷. 高原地区藏族中老年牙周炎与认知功能障碍相关性研究 [D]. 西宁: 青海大学, 2019.
- ZHAO T T. Correlation between periodontitis and cognitive dysfunction in Tibetan middle-aged and elderly people in plateau area [D]. Xining: Qinghai University, 2019.
- [79] CANO SIERRA J D, MESTRA C F, NIETO J C G, et al. Teeth infection may "shunt" through Fontan in high-altitude conditions [J]. Ann Transl Med, 2018, 6(7): 118.